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Sir:

Transmitted herewith for filing is a PROVISIONAL APPLICATION of

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for **NEW PEPTIDES**. The application comprises a **20**-page specification and **0** sheets of drawings.

Accompanying this application for filing is:

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New Peptides

Technical field and Background of the Invention

The present invention relates to synthetic peptides as well as to proteins related to these peptides and to pharmaceutical compositions comprising the peptides/proteins for the diagnosis and treatment of fertility-relevant and/or pregnancy-relevant autoantibodies.

Autoimmune diseases are disorders in which the immune system erroneously produces autoantibodies characterized by their binding to an endogenous antigen, with subsequent partial or complete loss of function of the endogenous antigen. Loss of function and other pathogenetic events are due to a number of effects, which can be associated with the binding of an antibody to an endogenous antigen:

Functionally relevant parts of the self-antigen (e.g. active enzymatic sites, binding areas, etc.) can be blocked by the antibody

The turnover of the protein can be increased and the active concentration be decreased

Antigen-Antibody complexes can be deposited and trigger pathogenetic pathways.

Compartmentalisation of the antigen can be influenced

Bound antibody can trigger a number of responses in the immune system, which secondarily influence the antigen itself or other parts of the body . Other autoantibody-mediated mechanisms might be possible, too. In addition, cell-mediated immune responses can contribute to the overall pattern of symptoms observed in a given autoimmune disease.

Several connective tissue disorders including vascular diseases such as vasculitis, systemic lupus erythematosus (SLE) and polymyositis, neurologic diseases such as multiple sclerosis and myasthenia gravis, and hematologic diseases such as idiopathic thrombocytopenic purpura (ITP) and anti-phospholipid syndrome (APS) seem to be caused by an autoimmune reaction. For some of these disorders, possible self antigens have been identified and/or pathogenic autoantibodies have been identified and isolated.

However, no specific drugs exist nowadays for the treatment of autoimmune diseases and patients are treated with general anti-inflammatory drugs such as corticosteroids

and/or symptom-specific drugs like anticoagulants (McIntyre et al. 2003) in the case of coagulation disorders due to autoantibodies.

Moreover, for many of the fertility-relevant autoantibody syndromes, such as SLE and APS, the antigens/antibodies causing the reproductive problems are not identified and the correlation between the general diagnostic detection of e.g. APS-autoantibodies and severity of reproductive symptoms is poor (Beer et al. 1998; Bermas et al. 1996a; Bermas et al. 1996b, 1996c; Bick and Baker 1999; Birdsall et al. 1996; Check 1998; Chilcott et al. 2000; Colaco and Male 1985). This is partly due to the fact, that most approaches to fertility-relevant autoantibodies did start with one of these broad polyclonal antibody syndromes like APS and identified prominent autoantibodies, which did not necessarily cause the reproductive problem, but were either representative for the whole syndrome or associated with another part of the syndrome.

It is an object of the invention to provide means for diagnosis of autoantibodies and specific treatment of autoantibody-related symptoms, which are associated with and/or cause reproductive problems. The present invention relates to novel peptides and proteins related to these peptides, which are anti-idiotypic to fertility-relevant autoantibodies. Thereby, the invention overcomes the lack of fertility-specific diagnostic and/or therapeutic means in the field of reproductive autoimmunity.

Description of the Invention

General Description

Peptides, which are able to bind autoantibodies being present in the blood of patients with reproductive problems, are obtainable by the following method:

A large number of biotinylated combinatorial peptides was synthesized by solid phase synthesis

These peptides were tested against plasma probes of patients with well-defined clinical symptomatology as well as against control samples

Peptides with specific recognition of patient samples (recognition defined by increased binding of human antibodies to the peptide as compared to the control samples) were tested in a larger population of samples.

Sequence variations of each positive peptide were tested to optimise a positive sequence.

Peptide sequences were used to check databases for sequence-related proteins harbouring the autoantibody target

Surprisingly, so far unknown peptides could be identified, which matched on fertility-relevant proteins.

Detailed Description

More than thousand random 12-mer peptide sequences were synthesized on an automated synthesis roboter and purified on an automated LC/MS-System, which enables the purification and analysis in a one-step approach. All sequences were foreseen with an additional terminal Glycine residue coupled to a biotin moleclule. Per peptide, an amount of 5-10mg – the final yield per specific peptide depending on the losses due to purification and problematic synthesis – were synthesized. A typical synthesis protocol is given as an example of the invention.

Plasma samples were obtained from patients with clinically well-documented reproductive problems. Samples from healthy female blood donors were used as reference.

Non-immunological causes (anatomical variations, endocrine problems, fertility problem of the male, etc.) of the respective reproductive disorders present in a given patient, were excluded. Moreover, all plasma samples were checked for the presence of anti-phospholipid antibodies. APS is an autoimmune syndrome, which is frequently associated with coagulation disorders, but also with fertility problems. Samples with anti-phospholipid antibody titres above the clinically relevant threshold were excluded from the first step of the peptide screen. Plasma samples were grouped according to the most common reproductive disorders:

Unexplained Infertility, in patients with repetitive (at least 2 times) failure in conventional in vitro-fertilisation procedures.

Habitual abortion, characterized by at least 3 consecutive abortions during the 1st trimester of pregnancy

Late pregnancy problems, this group being heterogenous and composed of plasma samples from patients with preeclampsia and/or severe intrauterine growth restriction of the fetus without maternal preeclamptic complications.

In an ELISA-protocol, the serum and plasma samples were tested against peptides from the available peptide pool. Basically, the ELISA was designed to detect antibodies being present in the patients blood, which bind to the peptides presented

to the sample. Technically, the ELISA followed the protocol given in Example 2 and comprised the following basic steps:

Binding of biotinylated peptides on streptavidin-coated Microtitre (96-well) plates.

Washing and Blocking of the wells

Incubation of the wells and appropriate control wells (without peptide) with the serum samples

Washing

Incubation with a suitable horseradish-peroxidase-labelled secondary antibody

Chromogenic detection and photometric read-out of the results.

In a first step, all peptides were tested against a reduced set of samples including pooled sample preparations in order to identify promising candidate peptides for lage scale analysis. In a second step, promising candidate peptides were tested against all samples to confirm the results obtained in the 1st screen. Using such ELISA-based protocols, it is easily possible to examine large numbers of blood samples and to test peptides against such samples. We could identify the following peptides, which showed clearly increased binding of patients antibodies as compared to control antibodies:

The Peptide 1 VYKSPNAYTLFS

The Peptide 2 RPEPQGAYLEQG

The Peptide 3 NSSYSPSLLESG

The Peptide 4 DQYIQQAHRSHI

The Peptide 5 QGLPAPQSYSRI

The Peptide 6 KQASNLTDMHYP

The Peptide 7 AQPNWTSLRSLP

The Peptide 8 HVNPHLHVHAWD

Based on these peptides, it is easily possible to deduce sequence variants, which also have the ability to bind antibodies occurring in the tested samples. For the skilled person in the field it is evident that conservative exchange of amino acids in the abovementioned peptide sequences will also lead to binding peptides, if the exchange

does not affect the physicochemical or structural properties of the peptide sequence to a large extent.

Using the peptide sequences as search sequences in BLAST-algorithms as they are offered e.g. by the NCBI or EMBL Web Portals ended up with proteins, which harbour sequences similar or identical to the peptides 1-3.

Surprisingly good hits on proteins were obtained with the sequences 1 and 3:

Peptide Sequence 1 shows a very good correlation with a sequence occurring in Pregnancy-associated Plasma Protein A (PAPP-A).

Peptide Sequence 2 shows has homologies with another region in PAPP-A.

Peptide Sequence 3 shows high homology with the Protein ADAM-TS13 (A disintegrin and a metalloproteinase and thrombospondin-13).

Amino acids described in this invention can be of the naturally occurring L stereoisomer form as well as the enantiomeric D form. The one-letter code refers to the accepted standard polypeptide nomenclature, but can mean alternatively a D- or L-amino acid:

Code	amino acids
A	L-Alanine or D-Alanine
V	L-Valine or D-Valine
L	L-Leucine or D-Leucine
I	L-Isoleucine or D-Isoleucine
M	L-Methionine or D-Methionine
F	L-Phenylalanine or D-Phenylalanine
Y	L-Tyrosine or D-Tyrosine
W	L-Tryptophan or D-Tryptophan
H	L-Histidine or D-Histidine
S	L-Serine or D-Serine
T	L-Threonine or D-Threonine
C	L-Cysteine or D-Cysteine
N	L-Asparagine or D-Asparagine

Q	L-Glutamine or D-Glutamine
D	L-Aspartic acid or D-Aspartic acid
E	L-Glutamic acid or D-Glutamic acid
K	L-Lysine or D-Lysine
R	L-Arginine or D-Arginine
P	L-Proline or D-Proline
G	Glycine

Examples of the invention

Synthesis of peptides, example

General Comments:

Unless stated otherwise a washing step is conducted by adding the solvent to the resin, shaking the mixture, and removing the solvent by vacuum filtration. At all steps it must be ensured that each resin bead is immersed in the reaction solution.

Step 1: Loading of the resin with the first amino acid

1 g 2-chlorotrityl resin (1.0 -2.0 mmol/g capacity) is suspended in 8 ml dichloromethane (DCM), shaken for 5 minutes at room temperature, and the solvent is removed by vacuum filtration. A solution of 2 mmol of the FMoc protected amino acid and 5 mmol (0.850 ml) diisopropylethylamine (DIPEA) in 8 ml DCM is added and the reaction is shaken for 1 hour at room temperature. After removing the reaction solution the resin is washed three times with 20 ml dimethylformamide (DMF) each. 20 ml of a mixture of DCM/Methanol/DIPEA 80:15:5 (v/v/v) is added, shaken for 15 to 30 minutes, the solution removed, and this step is repeated once. The resin is washed four times with 20 ml DMF each. The FMoc group is removed by adding 20 ml of 25 vol-% piperidine in DMF, shaking for 3 minutes, removing the solvent, adding another 20 ml of 25 vol-% piperidine in DMF, shaking for 30 minutes, and removing the solution by vacuum filtration. The resin is washed six times with 20 ml DMF each. In this state the resin can be stored overnight. For this purpose it has to be washed two times with 20 ml DCM each, and dried *in vacuo*. Should a second amino acid be coupled the procedure can be directly continued at step two instead of washing with DCM.

Step 2: Coupling of the 2nd amino acid

In the case that the resin has been stored overnight it has to be swollen by filling the reaction vessel completely with DMF. After 20 minutes the DMF is removed.

A solution of 5 mmol of the FMoc protected amino acid, that will be introduced, 7.5 mmol 1-hydroxybenzotriazole (HOBt), and 1 ml o DIPEA in 20 ml DMF (eventually up to 30 ml in the case that the amino acid derivative is not dissolved completely) is added to the resin. The suspension is vortexed for 5 minutes and 5 mmol of benzotriazole-1-yl-oxy-trispyrrolidinophosphonium hexafluorophosphate (PyBOP) is added as a solid as well as another ml of DIPEA. After vortexing for 60 to 90 minutes the reaction solution is filtered off, and the resin is washed 6 times with 30 ml DMF each. The resin can be stored in this state (after washing twice with DCM and drying *in vacuo*).

Step 3: Coupling of further amino acids

Further amino acids are coupled by removing the FMoc group with 25% piperidine in DMF (as described above) and repeating Step 2.

Step 4: Cleaving the peptide off the polymer

The resin which is loaded with the FMoc deprotected peptide is washed 6 times with 20 ml DMF and twice with 20 ml DCM each. 40 ml 2,2,2-trifluoroethanol/DCM 2:8 (v:v) is added. The reaction mixture is shaken from time to time and otherwise left standing for 60 minutes. The resin is filtered off and the filtrate co-evapourated several times with DCM.

Step 5: Deprotecting the peptide

40 ml of a mixture of trifluoroacetic acid/water/triisopropylsilan (TIS) 95:5:5 (v/v/v) is added. In the case that the solution is still coloured yellow after about 1 minute several drops of TIS are added. The mixture is left standing for about 60 minutes. Afterwards the cleaving mixture is removed by coevapourating several times with DCM. The product is dissolved in water (eventually adding a minimal amount of methanol) and lyophilisated. The crude peptide is purified by preparative HPLC.

Example of an ELISA-Protocol

Materials used in the protocol

96well streptavidin plates from Steffens (Steffens GmbH, Heidelberg, Germany)

Biotinylated IgG from Reactolab SA (5mg/ml)

Anti-human IgA,IgG,IgM from Sigma A8794 (22mg/ml)

Shaking at all incubation and washing steps was done on a laboratory shaking platform at 150rpm

Dilute APL peptides from (dimethylsulfoxide) DMSO stock (10mg/ml) into **washing buffer** to 1µg/ml (for this dilution, pipette: 1µl in 10ml)

Dilute stock of Reactolab solution 1:1000 in washing buffer +2% Milk powder +1%FCS, then:

For wells A 1,2: pipette 100µl washing buffer +2% Milk powder +1%FCS into each well.

For wells A 3,4: pipette 4µl of this solution into 2ml washing buffer +2% Milk powder +1%FCS, then pipette 100µl of this solution into each well.

For wells A 5,6: pipette 10µl of this solution into 2ml washing buffer +2% Milk powder +1%FCS, then pipette 100µl of this solution into each well.

For wells A 7,8: pipette 20µl of this solution into 2ml washing buffer +2% Milk powder +1%FCS, then pipette 100µl of this solution into each well.

For wells A 9,10: pipette 30µl of this solution into 2ml washing buffer +2% Milk powder +1%FCS, then pipette 100µl of this solution into each well.

For wells A 11,12: pipette 40µl of this solution into 2ml washing buffer +2% Milk powder +1%FCS, then pipette 100µl of this solution into each well.

Step	Standard (Row A)	Peptide and LW (B-H)
1	Equilibrate all wells in A (calibration curve*) with 100µl washing buffer + 2% milk powder + 1% FCS (10 min., Room Temperature)	Equilibrate all wells (B-H) with 100µl washing buffer (10 min., Room Temperature)
2	Pipette 100µl of the appropriately diluted standard solution into the respective wells and incubate 1 hour at RT.	Pipette 100µl of the diluted APL peptides (100ng total amount) in the wells, last row (H, containing Blank Values) without peptide (only washing buffer), incubate 1 hour at RT
3	Wash 3x 5 min. with washing buffer.	
4	Add 100µl washing buffer per well, incubate 1h RT	Add 1% human serum or plasma diluted in blocking buffer, 100µl per well, incubate 1h RT
5	Wash 3x 10min. with washing buffer	
6	Add 100µl anti-hu IgA,G,M-HRP 1:10000 (Stock: 22mg/ml) diluted in blocking buffer to all wells, incubate 1 hour at RT	
7	Wash 3x 5 min. with washing buffer	
8	Add 100µl OPD sunstrate to each well: 1 tablet OPD (20mg) in 33ml Citrate-phosphate buffer + 17µl 30% H_2O_2 (suffices for 3 plates)	
9	Incubate OPD for 10min and stop reaction with 100µl 1N HCL to each well	
10	Measure at measuring WL: 492 with reference WL: 620 in ELISA reader	

Washing buffer:

10mMTris,pH7,5 (10ml 1M Tris,pH7,5)

2,1% NaCl NaCl (21g)

2mM EDTA (4ml 0,5M EDTA)

1ml TritonX-100

ad aq. dest. to 1l

Citrate-Phosphate buffer (pH 5,0):

Citric acid.1 H_2O 7,3g

$Na_2HPO_4.2\ H_2O$ 11,86g

Ad aq.dest. to 1l

Blocking buffer:

Washing buffer + 2% milk powder **Plate outline**

		1	2	3	4	5	6	7	8	9	10	11	12
Standards	A	0	0	10ng	10ng	25ng	25ng	50ng	50ng	75ng	75ng	100ng	100ng
Peptides	B	Pep1	Pep1	Pep1	Pep1	Pep1	Pep1	Pep1	Pep1	Pep1	Pep1	Pep1	Pep1
	C	Pep2	Pep2	Pep2	Pep2	Pep2	Pep2	Pep2	Pep2	Pep2	Pep2	Pep2	Pep2
	D	Pep3	Pep3	Pep3	Pep3	Pep3	Pep3	Pep3	Pep3	Pep3	Pep3	Pep3	Pep3
	E	Pep4	Pep4	Pep4	Pep4	Pep4	Pep4	Pep4	Pep4	Pep4	Pep4	Pep4	Pep4
	F	Pep5	Pep5	Pep5	Pep5	Pep5	Pep5	Pep5	Pep5	Pep5	Pep5	Pep5	Pep5
	G	Pep6	Pep6	Pep6	Pep6	Pep6	Pep6	Pep6	Pep6	Pep6	Pep6	Pep6	Pep6
Blanks	H	ØPep	Ø	Ø	Ø	Ø	Ø	Ø	Ø	Ø	Ø	Ø	Ø
		Serum 1	Serum 2	Serum 3	Serum 4	Serum 5	Serum 6	Serum 7	Serum 8	Serum 9	Serum 10	Serum 11	Serum 12

Diagnostic potency of the peptides

In the following tables, the diagnostic potency of the peptides for pregnancy complications and fertility diagnostics is illustrated exemplary in tables. The groups of patients, which were analysed comprise

Apparently healthy female blood donors

Patients presenting the first time for ivF in an ivF-facility. These patients did not show any endocrine or morphological reason for infertility. There was no male infertility involved in the fertility problem of the couple. No other autoantibody diseases such as aPL-syndrome or Lupus erythematodes were known or detected.

Patients after repetitive ivF-failure (two cycles without pregnancy). Not autoimmune disease known and nor endocrine, morphological or male reasons known for infertility.

Patients with an anamnesis of habitual abortion, wherein these patients had at least two consecutive abortions in the 1st trimester of pregnancy.

Patients with an anamnesis of preeclampsia in at least one preceding pregnancy.

For optimal diagnostic sensitivity and specificity, the peptides can be combined to a small panel of diagnostic peptides. Abbreviations are as follows: MW (Mean); CI (confidence interval) C-off (cutoff value, defined as 1.1* (MW+CI)).

The Peptide shown in Table 1 exemplifies a peptide with a strong diagnostic profile in pregnancy complications, while infertility is detected much less sensitive.

Table 1 Peptide: DQYIQQAHRSHI	Female donors, Blood Bank	IvF	Repetitive ivF-failure	Habitual Abortion	Preeclampsia	
MW	6,87	8,72	10,11	17,64	18,44	
SD	7,24	13,31	8,70	14,65	9,29	
# of cases N	33	15	25	17	14	
99%CI	3,25	8,85	4,48	9,15	6,40	
MW+CI	10,12	17,57	14,59	26,79	24,84	
MW-CI	3,62	-0,13	5,63	8,49	12,04	
N>C-off	10	3	13	11	12	Number of cases above the Cut-OFF-level
Sensitivity	0,30	0,20	0,52	0,65	0,86	Fraction of those with the disease correctly identified by the test
Specificity	0,70	0,80	0,48	0,35	0,14	Fraction of those without the disease correctly identified by the test

The peptide shown in table 2 has profile, which shows no strong preference of one of the patient groups, but gives a relatively constant, relatively low sensitivity profile through all groups.

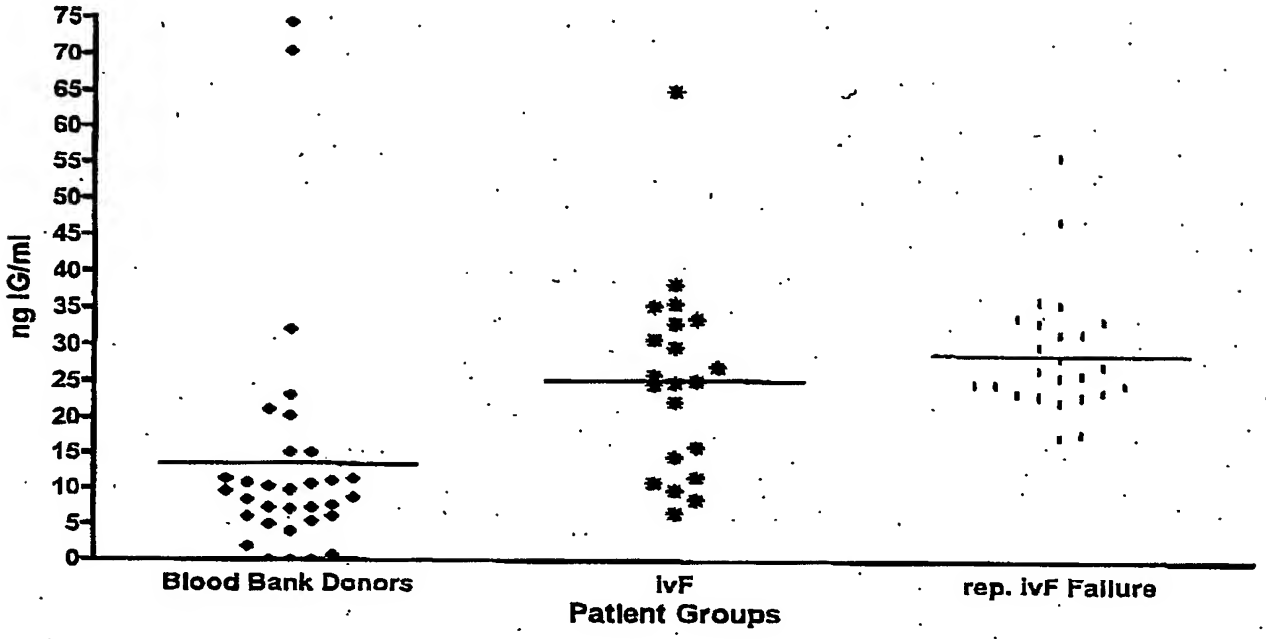
Table 2 Peptide: KQASNLTDMHYP	Female donors, Blood Bank	IvF	Repetitive ivF-failure	Habitual Abortion	Preeclampsia	
MW	2,57	4,95	7,87	11,52	8,53	
SD	3,45	3,84	10,69	12,25	10,80	
# of cases N	33	15	25	17	14	
99%CI	1,55	2,55	5,51	7,65	7,44	
MW+CI	4,11	7,50	13,38	19,17	15,97	
MW-CI	1,02	2,39	2,36	3,87	1,09	
N>C-off	10	8	12	10	6	Number of cases above the Cut-OFF-level
Sensitivity	0,30	0,53	0,48	0,59	0,43	Fraction of those with the disease correctly identified by the test
Specificity	0,70	0,47	0,52	0,41	0,57	Fraction of those without the disease correctly identified by the test

The peptide in table 3 shows a strong profile in the field of infertile patients, while no diagnostic potency is present in the field of pregnancy complications (habitual abortion and preeclampsia).

Table 3 Peptide: VYKSPNAYTLFS	Female donors, Blood Bank	IvF	Repetitive ivF-failure	Habitual Abortion	Preeclampsia	
[redacted]	17,23	27,55	31,55	7,82	7,19	
[redacted]	11,08	6,98	14,47	7,98	8,33	
[redacted]	33	14	25	51	42	
[redacted]	4,97	4,80	7,45	2,88	3,31	
[redacted]	22,20	32,35	39,00	10,70	10,50	
[redacted]	12,27	22,74	24,10	4,94	3,88	
N>C-off	11	12	21	4	4	Number of cases above the Cut-OFF-level
Sensitivity	0,33	0,86	0,84	0,08	0,10	Fraction of those with the disease correctly identified by the test
Specificity	0,67	0,14	0,16	0,92	0,90	Fraction of those without the disease correctly identified by the test

These results are also illustrated graphically in the following figure 1:

Figure 1: VYKSPNAYTLFS



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Claims

1. A method of assessing *ex vivo* the titre of fertility-relevant autoantibodies of a mammalian female, comprising the steps:
   - presentation of a molecule which binds to an idiotype of the fertility-relevant autoantibody, and
   - determining the amount of autoantibody being bound by said molecule from a sample of a body fluid of said mammalian female.
2. The method as claimed in claim 1, wherein said mammalian female is a human female.
3. The method as claimed in claims 1-2, wherein said body fluid is blood, serum, blood plasma or saliva.
4. The method as claimed in claims 1-3, wherein said molecule capable of binding to the idiotype of the fertility-relevant autoantibody is an oligo- or polypeptide.
5. The method as claimed in claims 1-4, wherein the oligopeptide is a peptide consisting of less than 40, most preferably less than 20 amino acids.
6. The method as claimed in claims 1-5, wherein the polypeptide is a naturally occurring protein, either being isolated from its natural source or being expressed in a recombinant host system, the latter thereby including modifications of the sequence by addition of sequence TAG's to facilitate purification or detection of the protein.
7. The method as claimed in claims 1-6, wherein the oligopeptides according to claim 5 are being part of the sequence of the polypeptide according to claim 6 meaning that the sequence of the oligopeptide is occurring in the sequence of the polypeptide.
8. The method as claimed in claims 1-7, wherein the oligopeptide is selected from one of the sequences (SEQ ID No. 1-8)

   VYKSPNAYTLFS (Seq ID No. 1)

   RPEPQGAYLEQG (Seq ID No. 2)

   NSSYSPSLLESG (Seq ID No. 3)

DQYIQQAHRSHI (Seq ID No. 4)

QGLPAPQSYSRI (Seq ID No. 5)

KQASNLTDMHYP (Seq ID No. 6)

AQPNWTSLRSLP (Seq ID No. 7)

HVNPHLHVHAWD (Seq ID No. 8)

or Retro-Inverso Derivatives of these peptide sequences.

9. The method as claimed in claims 1-8, wherein the amino acid sequence of the polypeptides mentioned in claim 8 is selected from the sequences of pregnancy-associated plasma protein A (PAPP-A; gi:38045915) or from ADAM-TS 13 (gi:21265049).

10. Oligopeptides having the sequences SEQ ID No. 1-8 or retro-inverso derivatives of these peptides, wherein the specific oligopeptide sequence can be extended at the amino-terminus and/or the carboxy-terminus by either up to 5 amino acids per terminus and/or be derivatized for detection by modifications including biotinylation, labelling by fluorochromes or radiolabelling.

11. The method as claimed in claims 1-9, wherein fertility-relevant autoantibodies being bound to the anti-idiotypic molecule are detected by an ELISA protocol using secondary antibodies being labelled with a detectable moiety such as an enzyme, a fluorochrome or biotin.

12. A diagnostic kit providing all reagents, standards, controls and accessories necessary to perform a method according to claims 1-8 and 10.

13. A pharmaceutical preparation containing an active ingredient according to claim 8 or a polypeptide having the sequence of pregnancy-associated plasma protein A (PAPP-A; gi:38045915) or from ADAM-TS 13 (gi:21265049).

Abstract

A method of assessing the titre of fertility-relevant autoantibodies of a mammalian female, wherein said method comprises:

- the presentation of a defined single molecule capable of binding to the idiotype of the fertility-relevant autoantibody, and
- determining the amount of autoantibody being bound by said molecule from a sample of a body fluid of said mammalian female.